

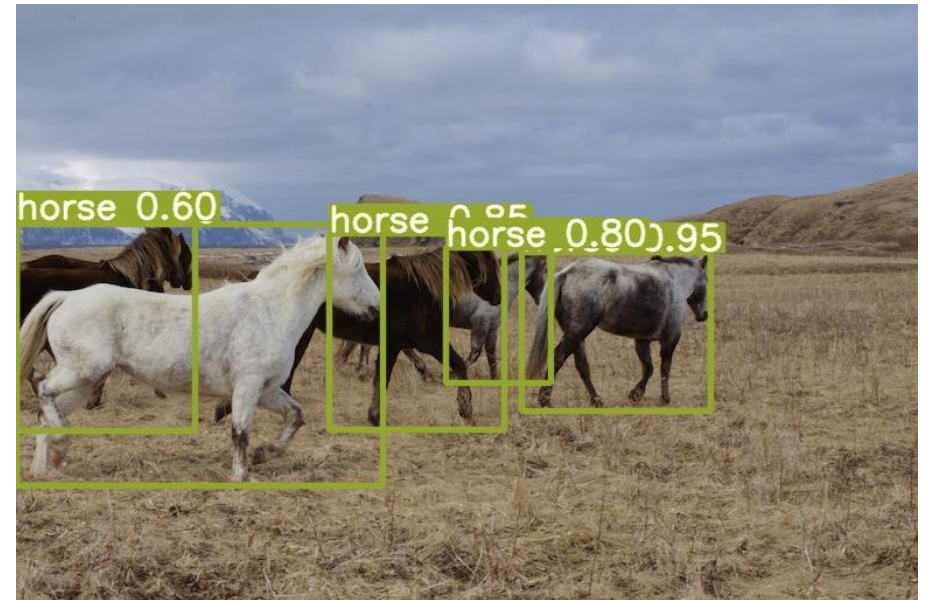
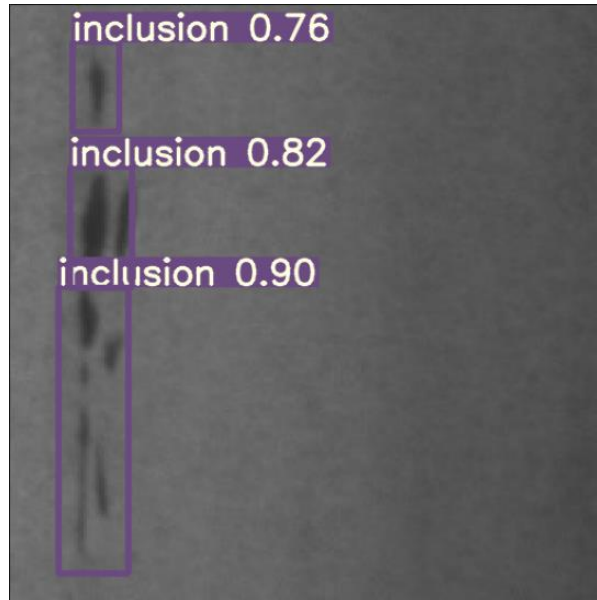
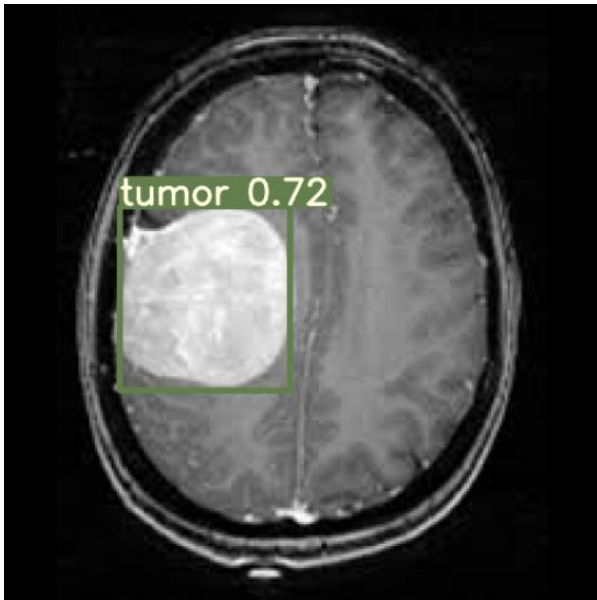
# Image-Object-Detection-PyTorch- YOLOR-GPL-Jupyter

One of Taiwan's proud recent new works, YOLOR, the most powerful object detection algorithm last year, greatly reduces the amount of computation, and increases the speed without reducing the accuracy. We organized the code so that we can use JupyterLab to perform the training and inference steps in sequence, which is easier to use, and produced an instruction slideshow.

Version 20230223

# Applications

- The YOLOR solution can be applied to factory defect detection, medical image analysis, biological image analysis, industrial safety image analysis, mask image analysis, etc.



# How to use

The main process is:

Annotate images -> Prepare files for training -> Training -> Inference

🏠 / Jupyter-Image-Object-Detection-YOLOR-GPL-1

Name

📁 data

📁 src

📄 1\_annotation\_pascal\_voc\_xml.ipynb

📄 2\_convert\_yolo\_format.ipynb

📄 3\_prepare\_train\_val\_txt.ipynb

📄 4\_delete\_log.ipynb

📄 5\_train.ipynb

📄 6\_tensorboard.ipynb

📄 7\_inference\_image.ipynb

📄 8\_inference\_image\_folder\_1.ipynb

📄 9\_inference\_webcam.ipynb

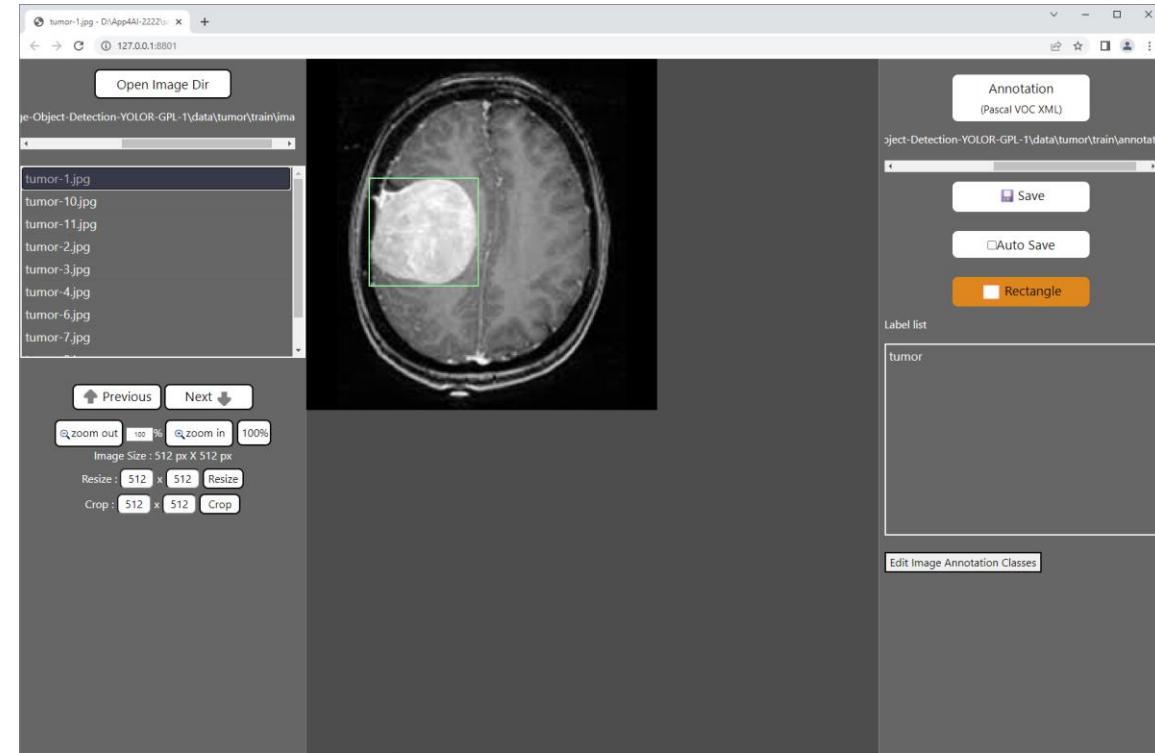
# 1\_annotation\_pascal\_voc\_xml.ipynb

Open the webpage for image annotation.

ipynb parameter:

- “port” is the port used by the webpage. If the port is occupied by the user, please change another port value by yourself.
- “dataset” is the dataset name
- “label\_folder” is the image of the train folder, it can also be changed to “val” to label the image of the val folder.

See Annotation.pdf for how to use annotation pages.

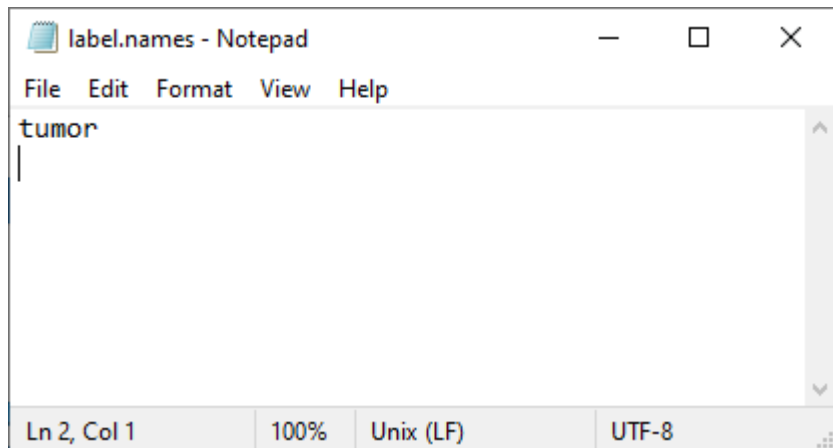


# 2\_convert\_yolo\_format.ipynb

Convert the voc xml label file to the yolo format. Before running, please confirm label.names under the label\_file path in #parameters and whether the content filled in the category is correct.

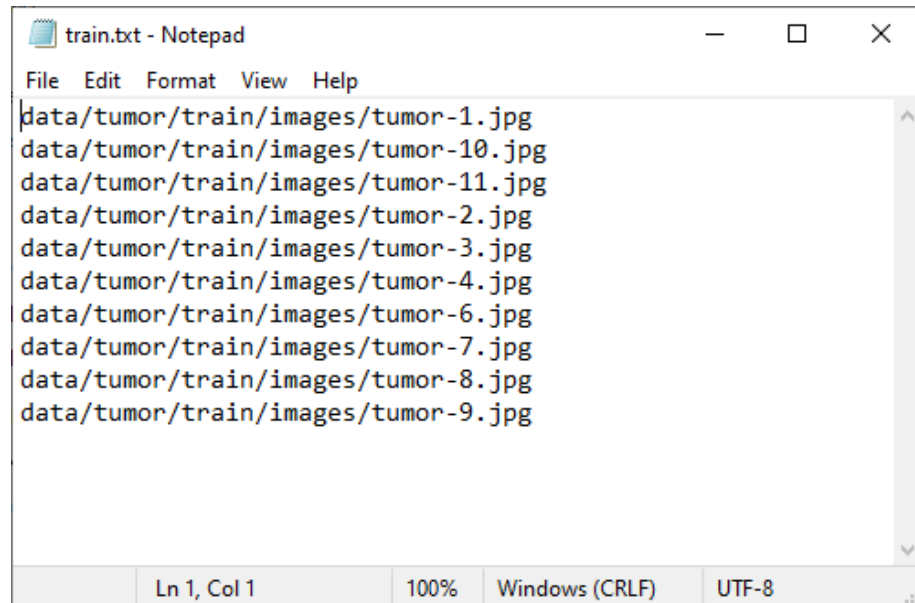
supplement:

The content of label.names is the category name without background.

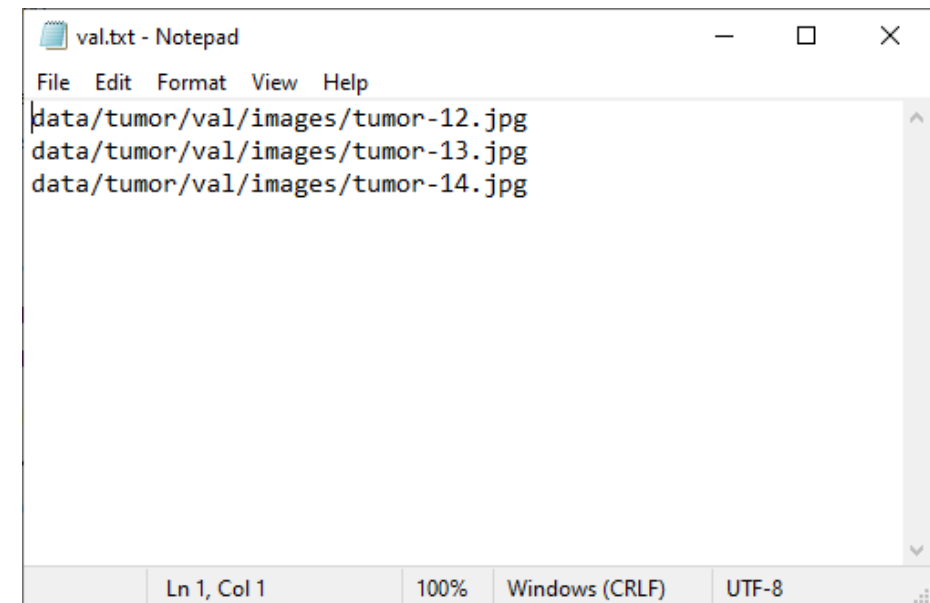


# 3\_prepare\_train\_val\_txt.ipynb

Generate training and validation image path files train.txt and val.txt .



```
train.txt - Notepad
File Edit Format View Help
data/tumor/train/images/tumor-1.jpg
data/tumor/train/images/tumor-10.jpg
data/tumor/train/images/tumor-11.jpg
data/tumor/train/images/tumor-2.jpg
data/tumor/train/images/tumor-3.jpg
data/tumor/train/images/tumor-4.jpg
data/tumor/train/images/tumor-6.jpg
data/tumor/train/images/tumor-7.jpg
data/tumor/train/images/tumor-8.jpg
data/tumor/train/images/tumor-9.jpg
Ln 1, Col 1 100% Windows (CRLF) UTF-8
```



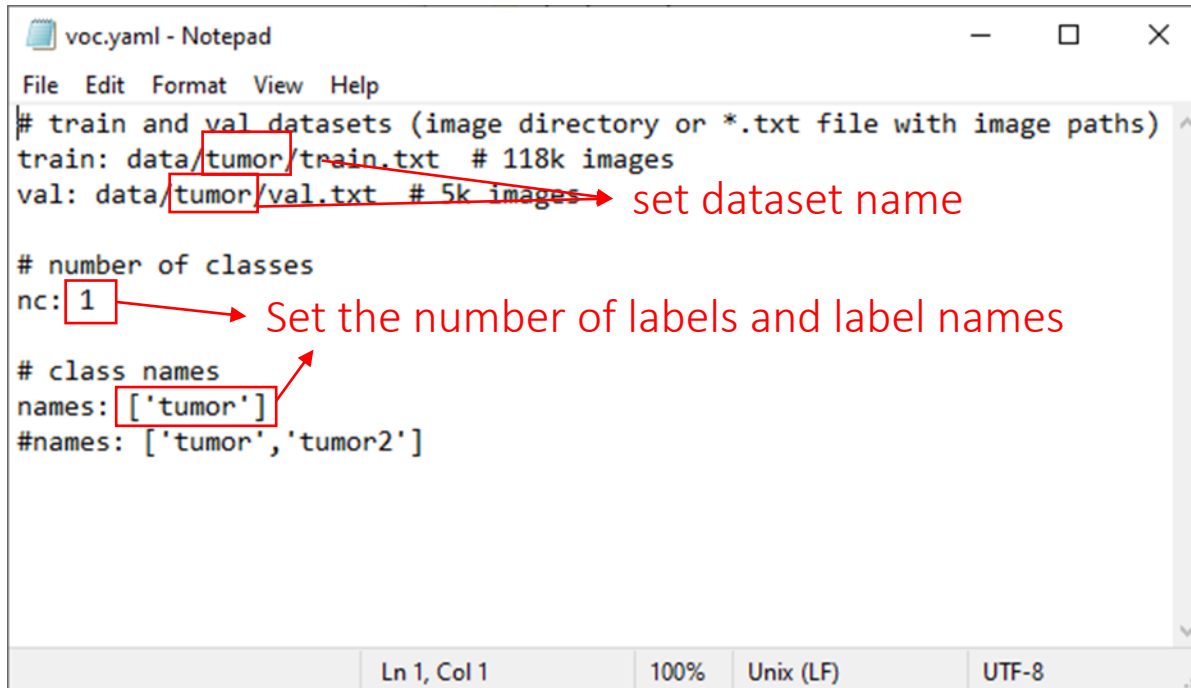
```
val.txt - Notepad
File Edit Format View Help
data/tumor/val/images/tumor-12.jpg
data/tumor/val/images/tumor-13.jpg
data/tumor/val/images/tumor-14.jpg
Ln 1, Col 1 100% Windows (CRLF) UTF-8
```

# 4\_delete\_log.ipynb

Delete the log files left over from previous training.

# Set the training voc.yaml related parameters

Set the content of the voc.yaml file in the dataset, set the name of the data set, the number of categories and the name.



```
File Edit Format View Help
# train and val datasets (image directory or *.txt file with image paths)
train: data/tumor/train.txt # 118k images
val: data/tumor/val.txt # 5k images
# number of classes
nc: 1
# class names
names: ['tumor']
#names: ['tumor', 'tumor2']
```

Annotations in the image:

- A red box highlights `data/tumor/` in both the `train` and `val` paths, with a red arrow pointing to the text "set dataset name".
- A red box highlights the value `1` in the `nc` field, with a red arrow pointing to the text "Set the number of labels and label names".
- A red box highlights the value `['tumor']` in the `names` field, with a red arrow pointing to the text "Set the number of labels and label names".

Footer information: Ln 1, Col 1 | 100% | Unix (LF) | UTF-8



# Set training yolor\_csp\_x.cfg related parameters

Set the content of the yolor\_csp\_x.cfg file in the dataset, set the number of categories.

The image displays three Notepad windows showing the configuration file `yolor_csp_x.cfg`. Red and blue annotations explain how to set the `filters` and `classes` parameters.

**Red Annotations:**

- Point to `filters=18` in the `[implicit_mul]` sections of the first and second windows.
- Text: "Set to (number of categories + 5) x 3 example: Suppose there is a label to train So  $(1 + 5) \times 3 = 18$ "

**Blue Annotations:**

- Point to `classes=1` in the `[yolo]` sections of the second and third windows.
- Text: "Set the number of categories example: Suppose there is a label to train So fill in 1"

**Content of the yolor\_csp\_x.cfg files:**

```
File Edit Format View Help
filters=320

# 193
[implicit_add]
filters=640

# 194
[implicit_add]
filters=1280

# 195
[implicit_mul]
filters=18

# 196
[implicit_mul]
filters=18

# 197
[implicit_mul]
filters=18

# ===== Head ===== #

# YOLO-3

[route]
Ln 1, Col 1 100% Unix (LF) UTF-8
```

```
File Edit Format View Help
pad=1
filters=18
activation=linear

[control_channels]
from=195

[yolo]
mask = 0,1,2
anchors = 23, 24, 44, 44, 69, 70, 150, 90, 80,175, 200,203, 456,141, 152,450, 456,454
classes=1
num=9
```

```
File Edit Format View Help
pad=1
filters=18
activation=linear

[control_channels]
from=196

[yolo]
mask = 3,4,5
anchors = 23, 24, 44, 44, 69, 70, 150, 90, 80,175, 200,203, 456,141, 152,450, 456,454
classes=1
num=9
Ln 1, Col 1 100% Unix (LF) UTF-8
```

# 5\_train.ipynb

Start training.

ipynb parameter:

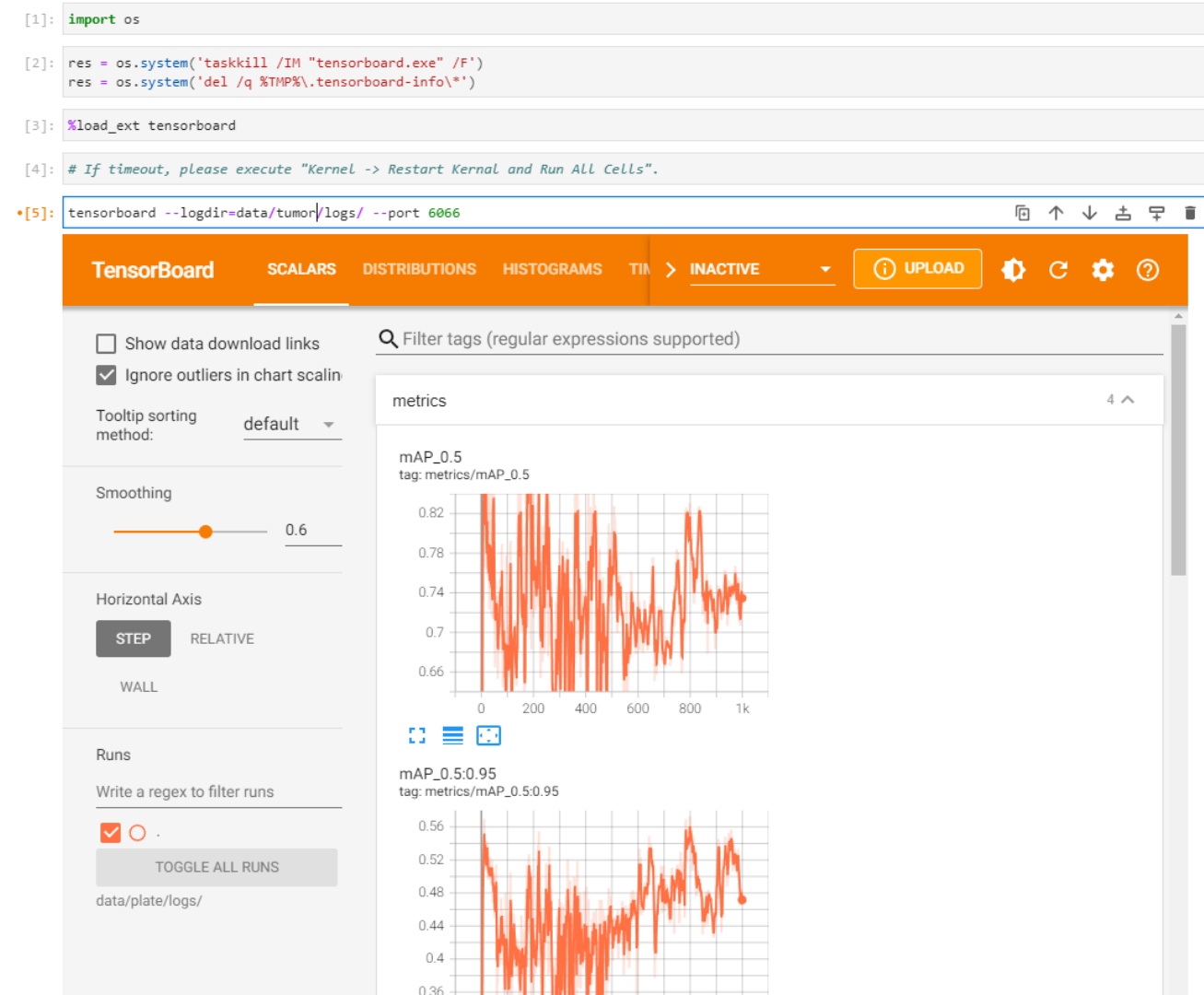
- dataset is the dataset name.
- weights\_file is the pretrained model path used, None means not to use the pretrained model for training.
- devices is the GPU id used.
- epochs is the number of training epochs.

```
Scanning labels data\tumor\train\labels.cache3 (10 found, 0 missing, 0 empty, 0 duplicate, for 10 images): 10it [00:00, 27.27it/s]
Scanning labels data\tumor\val\labels.cache3 (3 found, 0 missing, 0 empty, 0 duplicate, for 3 images): 3it [00:00, ?it/s]
Image sizes 512 train, 512 test
Using 1 dataloader workers
Logging results to data\tumor\model
Starting training for 1000 epochs...
```

Epoch	gpu_mem	box	obj	cls	total	targets	img_size
0/999	5.98G	0.03105	0.01745	0	0.0485	5	512: 100% █  2/2 [00:19<00:00, 9.63s/it]
1/999	7.39G	0.02041	0.008021	0	0.02843	1	512: 100% █  2/2 [00:00<00:00, 2.78it/s]
2/999	7.39G	0.01482	0.008739	0	0.02356	0	512: 100% █  2/2 [00:00<00:00, 2.37it/s]
3/999	7.39G	0.02104	0.008882	0	0.02992	2	512: 100% █  2/2 [00:00<00:00, 2.43it/s]
	Class	Images	Targets	P	R	mAP@.5	mAP@.5:.95: 0%   0/1 [00:00<?, ?itD:\App4AI-2222\gpu\pyth
	all	3	3	0.439	1	0.913	0.73
4/999	7.23G	0.02456	0.01079	0	0.03535	3	512: 100% █  2/2 [00:00<00:00, 2.46it/s]
	Class	Images	Targets	P	R	mAP@.5	mAP@.5:.95: 100% █  1/1 [00:00<00:00,
	all	3	3	0.441	1	0.913	0.73
5/999	7.24G	0.02236	0.01873	0	0.04109	2	512: 100% █  2/2 [00:00<00:00, 2.36it/s]
	Class	Images	Targets	P	R	mAP@.5	mAP@.5:.95: 100% █  1/1 [00:00<00:00,
	all	3	3	0.386	1	0.913	0.73
6/999	7.24G	0.02884	0.009161	0	0.038	2	512: 100% █  2/2 [00:00<00:00, 2.45it/s]
	Class	Images	Targets	P	R	mAP@.5	mAP@.5:.95: 100% █  1/1 [00:00<00:00,
	all	3	3	0.388	1	0.913	0.73

# 6\_tensorboard.ipynb

You can view the training loss curve and other related information through TensorBoard.



# 7\_inference\_image.ipynb

Infer a single image.

ipynb parameter:

- dataset is the dataset name.
- source is the inferred image path.
- weights\_file is the inference model path.

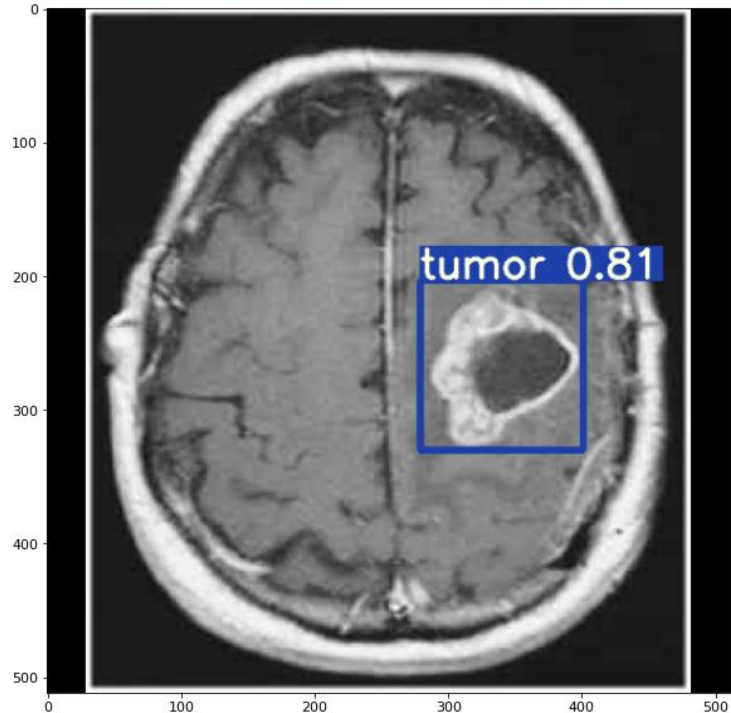
```
[4]: dataset = "tumor"
source = "data/%s/test/images/tumor-10.jpg" %(dataset)
image_size = 512
yaml_file = "data/%s/label.names"%(dataset)
cfg_file = "data/%s/yolor_csp_x.cfg"%(dataset)
weights_file = "data/%s/model/best.pt" %(dataset)
device = "0"
threshold = 0.6

[5]: %run src/detect.py --source $source --img-size $image_size --names $yaml_file --cfg $cfg_file --weights $weights_file --conf $threshold --device $device
```

Namespace(weights=['data/tumor/model/best.pt'], source='data/tumor/test/images/tumor-10.jpg', output='inference/output', img\_size=512, conf\_thresh=0.6, iou\_thres=0.5, device='0', view\_img=True, save\_txt=False, classes=None, agnostic\_nms=False, augment=False, update=False, cfg='data/tumor/yolor\_csp\_x.cfg', names='data/tumor/label.names', show\_rate=False, save\_img=False)

D:\App4AI-2222\gpu\python\lib\site-packages\torch\functional.py:568: UserWarning: torch.meshgrid: in an upcoming release, it will be required to pass the indexing argument. (Triggered internally at C:\actions-runner\work\pytorch\pytorch\buidler\windows\pytorch\aten\src\ATen\native\TensorShape.cpp:2228.)

```
return _VF.meshgrid(tensors, **kwargs) # type: ignore[attr-defined]
```



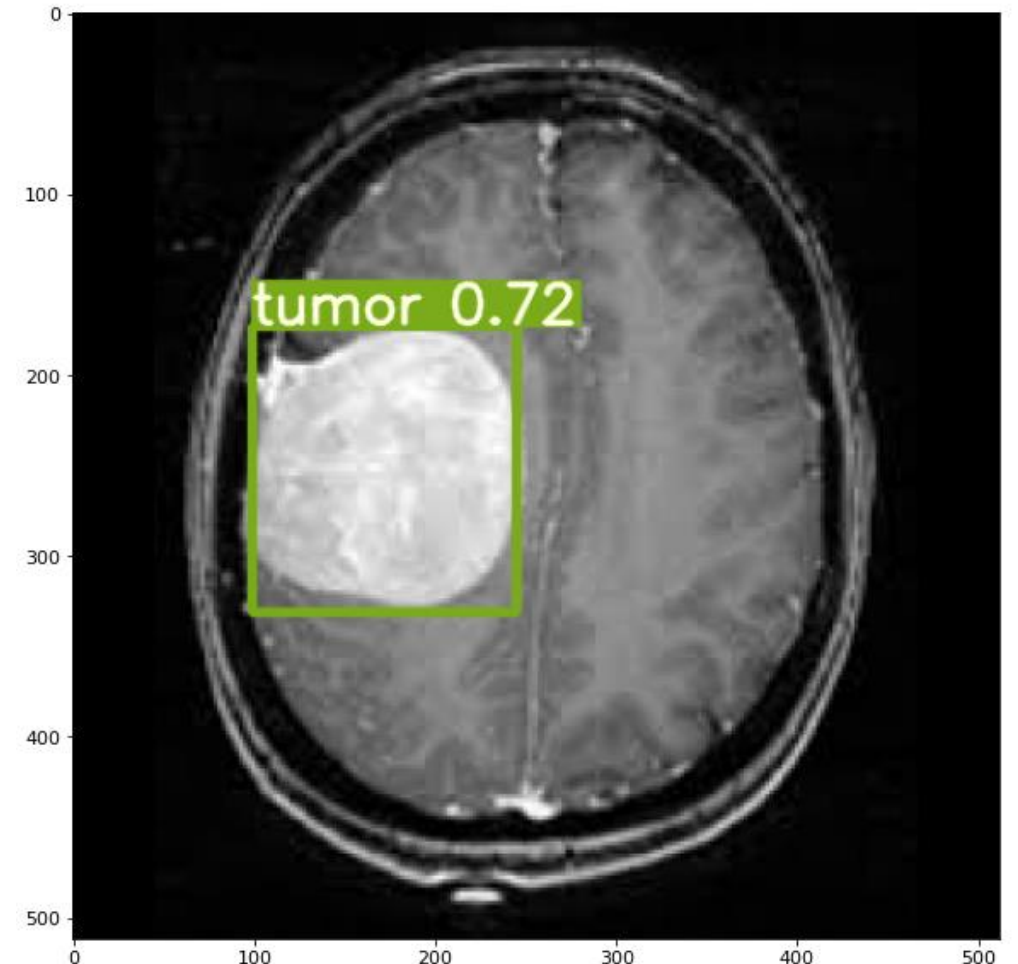
# 8\_inference\_image\_folder\_1.ipynb

Infer all images in the folder.

ipynb parameter:

- dataset is the dataset name.
- source is the inferred image path.
- weights\_file is the inference model path.

```
tumor-1  
tumor 0.716797  
Underkill Rate: 0(0.00%), Overkill Rate: 0(0.00%), Right Rate: 1(100.00%), Total: 1  
=====
```



# 9\_inference\_webcam.ipynb

Infer the image of the webcam. Press “q” on the display to turn the webcam off.

# Reference

- Please refer to the readme.txt in the SDK folder.
- LEADERG AppForAI: <https://www.leaderg.com/appforai-windows>
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